**John A. Lednicky, Ph.D.**

Associate Professor

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**Education**

Ph.D., Microbiology, University of Texas-Austin, 1991, ―Molecular-Genetic Analysis of the SV40 Upstream Promoter Elements

M.S., Microbiology, University of Missouri-Kansas City, 1984

B.S., Microbiology, University of Miami, Miami, Florida, 1978

**Certifications**

M(ASCP), Technologist in Microbiology, American Society of Clinical Pathologists, Certificate No. M001680, 1980

RM(NRCM), Registered Microbiologist, National Registry of Certified Microbiologists, certificate #2117

**Clearances**

Secret Security Clearance

Centers for Disease Control (CDC), Clearance for Select Agents, September 22, 2005

CDC PI Status, October 17, 2006

**Specialties**

Arboviruses: Detection of in clinical specimens or arthropods; isolation, identification, genetic analyses

*Avian influenza virus* H5N1 and other influenza viruses (human and animal): detection, isolation, and genetic analysis

Live agent bioaerosol inhalation studies with H5N1 and other viruses, and bacteria and fungi

Development of human and animal virus detection, isolation, and identification methodologies

Influenza virus assays (hemagglutination, hemagglutination inhibition, etc.)

Influenza virus vaccine safety and efficacy testing

Studies of respiratory viruses including adenoviruses, coronaviruses, rhinoviruses, hantaviruses and *SARS* coronavirus

Reverse genetics of RNA viruses: expression of viral genes and virus production

Polyomavirus (BKV, JCV, LPV, PyV, SV40) regulatory region structure and function

Molecular and classical diagnostic virology (human and animal)

Paramyxovirus detection, isolation, and genetic analysis (including *Canine distemper virus*, *Measles, Mumps, Human metapneumovirus, and* parainfluenza viruses) and immunogenicity of *Canine distemper virus*

Determination of whole genomic sequences of various DNA and RNA viruses

Diagnostic (clinical) microbiology (bacteriology, mycology, parasitology, and virology)

**Experience**

Associate Professor, Oct. 1, 2010 - Present

University of Florida, Gainesville

Environmental air sampling for the assessment of microbiology/virology air quality; arbovirology (detection, identification); identification of cervid viruses associated with hemorrhagic diseases in Florida; Development of air sampling methodologies for airborne viruses; Evaluation of inhalation nanoparticle toxicity and predisposition to influenza virus infection; swine model of NKT agonists for controlling influenza infections; Development of diagnostics tests and devices for *Zika virus*.

Senior Advisor, December 2009-Sept. 21, 2010

(Principal Scientist, July 2005-December 2009)

MRIGlobal, Kansas City, Missouri

Managed and provided technical oversight and guidance for programs involving molecular biology, virology, and microbiology; served as a corporate expert in the area of virology and molecular biology; developed and directed research programs; wrote and reviewed research proposals, technical reports, and presentations; and managed technical aspects, schedules, and budgets of programs. Recent projects included work with influenza viruses, alphaviruses, flaviviruses, *Bacillus anthracis*, SV40, *Canine distemper virus*, *Measles virus*, and algal culture. Established ferret model at MRI for intranasal and inhalation exposure studies with H5N1 and other influenza viruses. Established live agent bioaerosol capability for H5N1 and other respiratory pathogens with assistance of Richard Tuttle.

Assistant Professor, January 2001-June 2005

Department of Pathology, Loyola University Medical Center, Maywood, Illinois

Conducted molecular research on polyomaviruses and paramyxoviruses; established a new project on *Canine distemper virus* for the Conservation Medicine Center of Chicago that led to the isolation of significant new virulent isolates of the virus; fully sequenced five different *Canine distemper virus* genomes (information deposited at GenBank), the largest number by one as of July 2005; established collaborative projects on *Canine distemper virus;* established a *Canine distemper virus* testing service for the City of Chicago Animal Care and Control facility; helped the City of Chicago control a distemper outbreak in 2004; developed three projects for pathology residents and a medical student; lectured on various topics such as emerging viruses, paramyxoviruses, viral persistence, and on DNA structure and molecular biology principles; was involved with research rodent pathogen surveillance.

Research Assistant Professor, Department of Molecular Virology and Microbiology, 1997-2000

(Postdoctoral Fellow, Department of Molecular Virology and Microbiology, 1991-1996)

Baylor College of Medicine, Houston, Texas

Conducted molecular research on polyomavirus regulatory region structure and function, and on polyomavirus persistence and shedding; studied protein-protein interactions of polyomavirus tumor proteins and cellular proteins using the yeast two-hybrid system; developed molecular diagnostic techniques for detecting and identifying polyomaviruses; cloned and sequenced the most number of different SV40 strains by one individual as of July 2005 (sequences deposited in GenBank, molecular clones deposited at the American Type Culture Collection); developed SV40 phylogenic classification; cloned a new type of JCV genome; trained personnel in basic molecular biology and SV40 virology methods.

Research Associate, Department of Biochemistry, 1991

(Graduate Research Assistant, Department of Biochemistry, 1989-1991), University of Missouri-Columbia

Graduate Research and Teaching Assistant, Department of Microbiology, 1985-1989

(Graduate Teaching Assistant, Biology Laboratory, Department of Biology, 1984), University of Texas–Austin

Laboratory Instructor, 1983-1984, Rockhurst College, Kansas City, Missouri

Graduate Research Assistant, Department of Microbiology, 1981-1984, University of Missouri-Kansas City

Microbiology Technician, Clinical Microbiology Laboratory, 1979-1980, Med Labs, Inc., Denton, Texas

Microbiology Technician, Clinical Microbiology Laboratory, 1979, Bethany Medical Center, Kansas City, Kansas

**Professional Affiliations**

American Society for Microbiology

American Society for Virology

Pan American Society for Clinical Virology

Frontiers in Bioscience Society of Scientists (permanent member)

**Previous Memberships**

International Society for NeuroVirology

Wildlife Disease Association

American Association for the Advancement of Science

American Committee on Laboratory Animal Diseases

American Chemical Society

American Society of Clinical Pathologists

**Invited Member for Panels and Task Forces**

Panel-Audience Discussion 1: Issues related to the detection of SV40 DNA in human tissues. CBER-NCI-NICHD-NCID-NIP-NVPO Workshop, National Institutes of Health, Bethesda, Maryland (Jan. 27, 1997).

Invited participant: FDA-OVRR-CBER-sponsored SV40 PCR Working Group Meeting, National Institutes of Health, Bethesda, Maryland (July 1, 1997).

Panelist: International Myeloma Foundation Virus Symposium on SV40 and Human Cancer, Karolinska Institute, Stockholm, Sweden (Sept. 7, 1999).

Invited participant: Viruses and Human Cancer Workshop, sponsored by NCI; held at Bethesda Marriott Hotel, Bethesda, Maryland (March 12-13, 2001).

City of Chicago Commission on Animal Care and Control Task Force on *Canine distemper virus* (August 2004-June 2005).

**Committee Membership**

Applied Clinical Research Committee, Dept. of Pathology, Loyola (3/01 – 3/02)

Conservation Medicine Center of Chicago Research Committee (2/01 – 6/05)

Conservation Medicine Center of Chicago Steering Committee (4/01 – 6/05t)

Loyola University Medical Center Institutional Biohazard Committee (8/01 – 6/05)

Molecular Development Committee, Dept. of Pathology, Loyola (2/01 – 3/02)

Rodent User Committee, Loyola (7/16/02 – 6/05)

Awards Committee for the Medical School, Loyola (8/14/02 – 6/05)

BSI (Base Supplement Incentive) Compensation Committee, Loyola (2/18/05 – 6/05)

Leadership Council (previously Council of Principal Scientists), Midwest Research

Institute (7/05 – 9/10)

State of Missouri Laboratory Pandemic Influenza Preparedness Committee (1/19/06 –

2/07)

Institutional Biohazard Committee, Stowers Institute (5/06 – 9/10)

Steering Committee, Midwest Regional Center of Excellence [previously named

Scientific Advisory Committee, Midwest Regional Center of Excellence] (8/06 – 9/10)

Council of Principal Scientists Awards Committee, Midwest Research Institute (12/06)

Board of Governors, Council of Principal Scientists, Midwest Research Institute (1/07 – 2/09)

PHHP Research Day, Univ. FL-Gainesville (Ad Hoc Reviewer of Abstracts) (2/2011)

EGH Faculty search committee 2011

EGH Faculty search committee 2012

Research Committee, College of Public Health and Health Professions, UF (July 1, 2013 – June 30, 2015)

One Health Faculty search committee for Pre-Eminence Position (2013 – 2014)

EGH MPH and PhD student awards committee (April 2014)

EGH Search Committee for non tenure-track position in environmental toxicology (April 2014).

PHHP Faculty Council (July 2014 to present)

PHHP Chair Search for EGH (8/2015)

**Honors and Awards**

Dean’s List, University of Miami, 1975, 1976, 1978

Honor Society Membership, Phi Kappa Phi, 1986

McKinney Lewis Fellowship, University of Texas at Austin, 1987

Eklund Award for Excellence in Teaching, Department of Microbiology, University of Texas at Austin, 1988

Marquis Who’s Who in America

Delta Omega Honorary Society in Public Health, UF Beta Upsilon Chapter

**Publications and Papers**

Jackson, M., J. DeSena, **J. Lednicky,** B. McPherson, R. Haile, R. G. Garrison, and M. Rogolsky. 1983. Isolation and characterization of a bacteriophage factor that confers competence for genetic transformation to an exfoliative toxin-producing strain of *Staphylococcus aureus*. Infect. Immun. **39,** 939-947.

**Lednicky, J.**, and W. R. Folk. 1992. Two synthetic Sp1-binding sites functionally substitute for the 21-base-pair repeat region to activate simian virus 40 growth in CV-1 cells. J. Virol. **66,** 6379-6390.

**Lednicky, J. A.,** C. Wong, and J. S. Butel. 1995. Artificial modification of the viral regulatory region improves tissue culture growth of SV40 strain 776. Virus Research **35,** 143-153.

**Lednicky, J. A**., R. L. Garcea, D. J. Bersagel, and J. S. Butel. 1995. Natural simian virus 40 strains are present in human choroid plexus and ependymoma tumors. Virology **212**, 710-717.

Stewart, A. R., **J. A. Lednicky**, U. Benzick, M. J. Tevethia, and J. S. Butel. 1996. Identification of a variable region at the carboxy terminus of SV40 large T-antigen. Virology **221**, 355-361.

**Lednicky, J. A.,** S. Jafar, C. Wong, and J. S. Butel. 1997. High-fidelity PCR amplification of infectious copies of the complete simian virus 40 genome from plasmids and virus-infected cell lysates. Gene **184**, 189-195.

**Lednicky, J. A.**, and J. S. Butel. 1997. A coupled PCR and restriction digest method for the detection and analysis of the SV40 regulatory region in infected-cell lysates and clinical samples. J. Virol. Methods **64**, 1-9.

**Lednicky, J. A.,** and J. S. Butel. 1997. Tissue culture adaptation of natural isolates of SV40: changes occur in viral regulatory region but not in carboxy-terminal domain of large T-antigen. J. Gen. Virol. **78,** 1697-1705.

**Lednicky, J. A.,** A. R. Stewart, J. J. Jenkins III, M. J. Finegold, and J. S. Butel. 1997. SV40 DNA in human osteosarcomas shows sequence variation among T-antigen genes. Int. J. Can. **72,** 791-800.

Rubelj, I., Venable, S. F., **Lednicky, J.,** Butel, J. S., Bilyeu, T., Darlington, G., Surmacz, E., Campisi, J., and Pereira-Smith, O. 1997. Loss of T-antigen sequences allows SV40-transformed human cells to escape crisis and acquire the senescent phenotype. J. Gerontology **52A**, B229-234.

Stewart, A. R., **J. A. Lednicky**, and J. S. Butel. 1998. Sequence analyses of human tumor-associated SV40 DNAs and SV40 viral isolates from monkeys and humans. J. Neurovirol. **4**, 182-193.

**Lednicky, J. A.,** A. S. Arrington, A. R. Stewart, X. M. Dai, C. Wong, S. Jafar, M. Murphey-Corb, and J. S. Butel. 1998. Natural isolates of simian virus 40 from immunocompromised monkeys display extensive genetic heterogeneity: New implications for polyomavirus disease. J. Virol. **72**, 3980-3990.

Butel, J. S., A. S. Arrington, C. Wong, **J. A. Lednicky**, and M. J. Finegold. 1999. Molecular evidence of SV40 infections in children. J. Infectious Diseases  **180,** 884-887.

Arrington, A. S., J. A. Lednicky, and J. S. Butel. 2000. Molecular characterization of SV40 DNA in multiple samples from a human mesothelioma. Anticancer Research **20,** 879-884.

Strickler, H. D., et al. 2001. A multicenter evaluation of assays for detection of SV40 DNA and results in masked mesothelioma specimens. Cancer Epidemiol., Biomarkers, and Prevention **10,** 523-532.

Vilchez, R. A., **J. A. Lednicky**, S. J. Halvorson, Z. S. White, C. A. Kozinetz, and J. S. Butel. 2002. Detection of polyomavirus SV40 tumor antigen DNA in AIDS-related systemic Non-Hodgkin’s lymphoma. J. Acquir. Immune Defic. Syndro. **29,** 109-116.

**Lednicky, J. A.**, S. J. Halvorson, and J. S. Butel. 2002. Detection and DNA sequence analysis of the regulatory region of lymphotropic papovavirus in peripheral blood mononuclear cells of a simian immunodeficiency virus-infected Rhesus macaque with simian virus 40 disease. J. Clin. Microbiol. **40,** 1056-1059.

**Lednicky, J. A.**, R. A. Vilchez, W. A. Keitel, F. Visnegarwala, Z. S. White, C. A. Kozinetz, D. E. Lewis, and Janet S. Butel. 2003. Polyomavirus JCV excretion and genotype analysis in HIV-infected patients receiving highly active antiretroviral therapy. AIDS **17,** 801-807.

Ling, P. A., **J. A. Lednicky,** W. A. Keitel, David Poston, Z. S. White, R-S Peng, Z. Liu, S. K. Mehta, D. L. Pierson, C. M. Rooney, R. A. Vilchez, E. O’Brien Smith, and J. S. Butel. 2003. The dynamics of herpesvirus and polyomavirus reactivation and shedding in healthy adults: a 14-month longitudinal study. Journal of Infectious Diseases **187,** 1571 – 80.

**Lednicky, J. A.**, T. P. Meehan, M. J. Kinsel, J. Dubach, L. L. Hungerford, N. A. Sarich, K. E. Witecki, M. D. Braid, C.Pedrak, and C. M. Houde. 2004.Effective primary isolation of wild-type *Canine distemper virus* in MDCK, MV1 Lu and Vero cells without nucleotide sequence changes within the entire haemagglutinin protein gene and in subgenomic sections of the fusion and phospho protein genes. Journal of Virological Methods **118**, 147-157

Rubinas, T. C., R. B. Carey, M. C. Kampert, S. Alkan, and **J. A. Lednicky**. 2004. Fatal Hemorrhagic Pneumonia Concomitant with *Chlamydia pneumoniae* and *Parainfluenza virus 4* Infection. Archives of Pathology and Laboratory Medicine **128**, 640-644.

Wright, M.H., L. M. Cera, N. A. Sarich, and **J. A. Lednicky.** 2004. Reverse Transcription – Polymerase Chain Reaction Detection and Nucleic Acid Sequence Confirmation of Reovirus Infection in Laboratory Mice with Discordant Serologic Indirect Immunofluorescence Assay and Enzyme-Linked Immunosorbent Assay Results. Comparative Medicine **54**, 410 - 417.

Zdziarski, J.M., N. A. Sarich, K. E. Witecki,and **J. A. Lednicky.** 2004. Molecular Analysis of SV-40-CAL, a New Slow Growing SV-40 Strain from the Kidney of a Caged New World Monkey with Fatal Renal Disease. Virus Genes **29,** 183-190.

Forsman, Z. H., **J. A. Lednicky**, G. E. Fox, R. C. Willson, Z. S. White, S. J. Halvorson, C. Wong, A. M. Lewis, Jr, and J. S. Butel.2004. Phylogenetic analysis of polyomavirus simian virus 40 from monkeys and humans reveals genetic variation.J Virol. **78,** 9306-9316.

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[Cutrone, R](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Cutrone+R%22%5BAuthor%5D)., **J.** [**Lednicky**](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Lednicky+J%22%5BAuthor%5D), G. [Dunn](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Dunn+G%22%5BAuthor%5D), P. [Rizzo](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Rizzo+P%22%5BAuthor%5D), M. [Bocchetta](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Bocchetta+M%22%5BAuthor%5D), K. [Chumakov](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Chumakov+K%22%5BAuthor%5D), P. [Minor](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&cmd=Search&term=%22Minor+P%22%5BAuthor%5D), and M. Carbone M. 2005. Some oral poliovirus vaccines were contaminated with infectious SV40 after 1961. [Cancer Res.](javascript:AL_get(this,%20'jour',%20'Cancer%20Res.');) **65,**10273-10279.

Hamilton, S. B., D. E. Daniels, W. A. Sosna, E. R. Jeppesen, J. M. Owells, M. D. Halpern, K. S. McCurdy, J. O. Rayner, and **J. A. Lednicky**. 2010. [Gas-permeable ethylene bags for the small scale cultivation of highly pathogenic avian influenza H5N1 and other viruses in embryonated chicken eggs.](http://www.ncbi.nlm.nih.gov/pubmed/20109234?itool=EntrezSystem2.PEntrez.Pubmed.Pubmed_ResultsPanel.Pubmed_RVDocSum&ordinalpos=1) Virol J. **7:**e23.

**Lednicky, J. A.,** J. M.[Villanueva](http://www.ncbi.nlm.nih.gov/pubmed?term=%22Villanueva%20JM%22%5BAuthor%5D), S. A. [Burke](http://www.ncbi.nlm.nih.gov/pubmed?term=%22Burke%20SA%22%5BAuthor%5D), R. Shively, M. W. [Shaw](http://www.ncbi.nlm.nih.gov/pubmed?term=%22Shaw%20MW%22%5BAuthor%5D), D. E. [Daniels](http://www.ncbi.nlm.nih.gov/pubmed?term=%22Daniels%20DE%22%5BAuthor%5D), S. B. Hamilton, and R. O. [Donis](http://www.ncbi.nlm.nih.gov/pubmed?term=%22Donis%20RO%22%5BAuthor%5D). 2010. Validation of a Method for Preparing Influenza H5N1 Simulated Samples. Journal of Virological Methods **167:**125-131.

Tuttle, R. S., W. A. Sosna, D. E. Daniels, S. B. Hamilton, and **J. A. Lednicky.** 2010. [Design, assembly, and validation of a nose-only inhalation exposure system for studies of aerosolized viable influenza H5N1 virus in ferrets.](http://www.ncbi.nlm.nih.gov/pubmed/20573226) Virol J. 2010 **7:**e135.

**Lednicky, J. A.,** S. B. Hamilton, R. S, Tuttle, W. A. Sosna, D. E. Daniels, and D. E. Swayne. 2010. Ferrets develop fatal influenza after inhaling small particle aerosols of highly pathogenic avian influenza virus A/Vietnam/1203/2004 (H5N1) Virol. J. **7:**e231

**Lednicky, J. A.,** C. R. Croutch, S. J. Lawrence, S. B. Hamilton, D. E. Daniels, and A. B. Astroff. 2010. A Non-Lethal Young Domesticated Ferret (*Mustela putorius furo*) Model for Studying Pandemic *Influenza Virus* A/California/04/2009 (H1N1). Comparative Medicine. **60:** 364-368.

Hamilton, S. B., D. E. Wyatt, B. T. Wahlgren, M. K. O’Dowd, J. M. Morrissey,D. E. Daniels, and **J. A. Lednicky.** 2011. Higher titers of some H5N1 and recent human H1N1 and H3N2 influenza viruses in Mv1 Lu vs. MDCK cells. Virol. J. **8:**e66.

**Lednicky, J. A.**, T. B. Waltzek, M. D. Halpern, and S. B. Hamilton. 2012. [Comparative analysis of the full-length genome sequence of a clinical isolate of Human parainfluenza virus 4B](http://www.scientifica.com/aip/871201). Scientifica e871201.

**Lednicky JA**, Waltzek TB, McGeehan E, Loeb JC, Hamilton SB, Luetke MC. Isolation and genetic characterization of human coronavirus NL63 in primary human renal proximal tubular epithelial cells obtained from a commercial supplier, and confirmation of its replication in two different types of human primary kidney cells. Virol J. 2013 Jun 27;10:213.

**Lednicky JA,** Loeb JC. Detection and Isolation of Airborne Influenza A H3N2 Virus Using a Sioutas Personal Cascade Impactor Sampler. Influenza Res Treat. 2013;2013:656825.

**Lednicky JA,** Butel JS, Luetke MC, Loeb JC. Complete genomic sequence of a new Human polyomavirus 9 strain with an altered noncoding control region. Virus Genes. 2014 Dec;49(3):490-2.

Memish ZA, Almasri M, Assirri A, Al-Shangiti AM, Gray GC, **Lednicky JA,** Yezli S. Environmental sampling for respiratory pathogens in Jeddah airport during the 2013 Hajj season. Am J Infect Control. 2014 Dec;42(12):1266-9.

Sanpui P, Zheng X, Loeb JC, Bisesi JH Jr, Khan IA, Afrooz AR, Liu K, Badireddy AR, Wiesner MR, Ferguson PL, Saleh NB, **Lednicky JA,** Sabo-Attwood T. Single-walled carbon nanotubes increase pandemic influenza A H1N1 virus infectivity of lung epithelial cells. Part Fibre Toxicol. 2014 Dec 14;11:66.

Fennelly KP, Tribby MD, Wu C-Y, Heil GL, Radonovich LJ, Loeb JC, **Lednicky JA**. Collection and measurement of aerosols of viable influenza virus in liquid media in an Andersen cascade impactor. Virus Adaptation and Treatment. 2014 Dec; 7:1–9.

Sayler KA, Barbet AF, Chamberlain C, Clapp WL, Alleman R, Loeb JC, **Lednicky JA**. [Isolation of Tacaribe virus, a Caribbean arenavirus, from host-seeking Amblyomma americanum ticks in Florida.](https://www.ncbi.nlm.nih.gov/pubmed/25536075) PLoS One. 2014 Dec 23;9(12):e115769.

Perri MG, Peoples-Sheps M, Blue A, **Lednicky JA,** Prins C. Public health education at the University of Florida: synergism and educational innovation. Am J Public Health. 2015 Mar;105 Suppl 1:S83-7. doi: 10.2105/AJPH.2014.302414.

Iovine NM, Morris JG Jr, Fredenburg K, Rand K, Alnuaimat H, Lipori G, Brew J, **Lednicky JA.** [Severity of influenza A(H1N1) illness and emergence of D225G variant, 2013-14 influenza season, Florida, USA.](https://www.ncbi.nlm.nih.gov/pubmed/25811540) Emerg Infect Dis. 2015 Apr;21(4):664-7.

**Lednicky JA,** Iovine NM, Brew J, Loeb JC, Sugimoto JD, Rand KH, Morris JG. [Hemagglutinin Gene Clade 3C.2a Influenza A(H3N2) Viruses, Alachua County, Florida, USA, 2014-15.](https://www.ncbi.nlm.nih.gov/pubmed/26692074) Emerg Infect Dis. 2016 Jan;22(1):121-3.

Pan M, Eiguren-Fernandez A, Hsieh H, Afshar-Mohajer N, Hering SV, **Lednicky J**, Hugh Fan Z, Wu CY. Efficient collection of viable viral aerosol through laminar flow water-based condensational particle growth. J Appl Microbiol. 2016 Mar;120(3):805-15.

Khan E, Farooqi JQ, Barr KL, Prakoso D, Nasir A, Kanji A, Shakoor S, Malik FR, Hasan R, **Lednicky JA**, Long MT. Flaviviruses as a cause of undifferentiated fever in Sindh Province, Pakistan: A preliminary report. Front Public Health. 2016 Feb 16;4:8.

Artiaga BL, Yang G, Hackmann TJ, Liu Q, Richt JA, Salek-Ardakani S, Castleman WL, **Lednicky JA,** Driver JP. α-Galactosylceramide protects swine against influenza infection when administered as a vaccine adjuvant. Sci Rep. 2016 Mar 23;6:23593.

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Jiang X, Pan M, Hering SV, **Lednicky J**, Wu CY, Fan ZH. Use of RNA amplification and electrophoresis for studying virus aerosol collection efficiency and their comparison with plaque assays. Electrophoresis. 16 May 2016. elps.201600141.

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Elbadry M, **Lednicky J,** Cella E, Telisma T, Chavannes S, Loeb J, Ciccozzi M, Okech B, De Rochars VM, Salemi M, Morris JG Jr. Isolation of an Enterovirus D68 from blood from a child with pneumonia in rural Haiti: Close phylogenetic linkage with New York strain. Pediatr Infect Dis J. 2016 Sep;35(9):1048-50.

**Lednicky JA**, Bonny TS, Morris JG, Loeb JC. Complete Genome Sequence of Enterovirus D68 Detected in Classroom Air and on Environmental Surfaces. Genome Announc. 2016 Jun 16;4(3). pii: e00579-16. doi: 10.1128/genomeA.00579-16.

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Tania S. Bonny, Maohua Pan, Julia C. Loeb, Xiao Jiang, Arantzazu Eiguren-Fernandez, Susanne Hering, Z. Hugh Fan, Chang-Yu Wu, **John A. Lednicky**. Drifted Influenza A and B Viruses Collected by a Water-Based Condensation Growth Air Sampler in a Student Healthcare Center during an Influenza Outbreak. Genome Announc. 2017 Apr 13;5(15). pii: e00178-17. doi: 10.1128/genomeA.00178-17.

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**Lednicky, J. A.,** and A. B. Astroff. 2010. [Pandemic preparedness & the development of vaccines for H5N1 influenza at the Midwest Research Institute.](http://www.ncbi.nlm.nih.gov/pubmed/21207777) Mo Med. 107:298-301.

**Non-peer-reviewed article (aquaculture related)**

**Lednicky, J. A.** 2004. Bumblebee goby: Effective maintenance and breeding, and raising of fry. Aquarticles. Accessible at: http://aquarticles.com/articles/breeding/Lednicky\_Bumblebee\_Goby.html

**Technical newsletter**

Sue Denny, **John Lednicky**, and Ralph Horne. Avian Influenza (H5N1) Laboratory Fact Sheet. December 2006. Missouri Department of Health and Senior Services, State Public Health Laboratory.

# Dr. John Lednicky, Dr. Jonathan Rayner, Dr. David Franz. Resources for Information on the H1N1 Swine Influenza Virus and Recommendations for Vaccination. Sept 14, 2009. Presented by MRI to US Senator Sam Brownback (Kansas).

**Review Articles**

Butel, J. S., J. A. Lednicky, A. R. Stewart, R. L. Garcea, and M. J. Finegold, ―SV40 and human brain tumors, *J. Neurovirology*, 3 Suppl. 1, S78-79 (1997).

Butel, J. S., and J. A. Lednicky, ―The cell and molecular biology of SV40: review and assessment of implications for human infections and disease, *J. Nat. Can. Inst.*, 91, 119-134 (1999).

Lednicky, J. A., and J. S. Butel, ―Polyomaviruses and tumors: possible significance of association, *Frontiers in Bioscience*, 4, d153-164 (1999).

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Lednicky, J. A., ―Hantaviruses: A short review, *Archives of Pathology and Laboratory Medicine*, 127, 30-35 (2003).

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**Book Chapters**

Folk, W. R., W. J. Tang, M. Martin, J. Lednicky, S. Berger, and R. H. Adams. 1988. Polyomavirus sequences affecting the initiation of transcription and DNA replication, in: *Molecular Aspects of Papovaviruses,* Martinus Nijhoff Publishing, Boston (Yosef Aloni, ed.).

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Lednicky, J.A. and Wyatt, D. E. 2012. The Art of Animal Cell Culture for Virus Isolation. In: Tissue Culture, ed: InTech, ISBN 980-953-307-097-6, Zagreb, Croatia. <http://www.intechopen.com/articles/show/title/the-art-of-animal-cell-culture-for-virus-isolation>

**Recent Abstracts and Poster Presentations**

Modulation of influenza virus infectivity and activation of toll-like receptors by carbon nanomaterials. X. Zheng, P. Sanpui, **J. Lednicky**, J. Loeb, T. Sabo-Attwood. EPI Research Day, University of Florida, (February 13, 2013).

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| Anderson BD, Ma M, Xia Y, Wang T, Shu B, **Lednicky JA**, Gray GC. A One Health Approach for Studying Swine Influenza Virus Transmission in Pig Farms, China. International Conference on Emerging Infectious Diseases, Atlanta, Georgia, USA, August 24-26, 2015.  Maohua Pan, Arantzazu Eiguren-Fernandez, Nima Afshar-Mohajer, Susanne Hering, Chang-Yu Wu, **John Lednicky**, Hugh Fan, and Hsin Hsieh. A Novel Sampler for Viral Aerosols through Water-based Condensation Particle Growth. AAAR 34th Annual Conference in Minneapolis, MN, USA, October 12-16, 2015.  Kenneth H. Rand, M.D., Maura Pieretti PhD, Rodney Arcenas PhD, Stacy G. Beal, M.D., and **John Lednicky Ph**D. Semi-Quantitative Patient Data from a Multiplex Respiratory Viral Panel (RVP): Can We Learn Something from Population-Based Study? Association for Molecular Pathology (AMP) 2015 Annual Meeting, Nov. 5 – 7, Austin, Texas.  **John Lednicky**, Maohua Pan, Julia Loeb, Hsin Hsieh, Arantzazu Eiguren-Fernandez, Nima Afshar-Mohajer, Susanne Hering, Chang-Yu Wu, Hugh Fan. Highly Efficient Collection of Viable Influenza Virus A/Mexico/4108/2009 (pdmH1N1). AAAR 34th Annual Conference in Minneapolis, MN, USA, October 12-16, 2015.  Hoaran Hu, Chang-Yu Wu, Nima Afshar-Mohajer, **John A. Lednicky,** Z. Hugh Fan, and Alexander Theodore. Size amplification of virus aerosol by batch adiabatic-expansion for size intensification by condensation (BASIC). AAAR 34th Annual Conference in Minneapolis, MN, USA, October 12-16, 2015.  **John Lednicky**, Maohua Pan, Julia Loeb, Hsin Hsieh, Arantzazu Eiguren-Fernandez, Mohajer, Susanne Hering, Chang-Yu Wu, Hugh Fan, Nima Afsha-Mohajer. Collection of viable airborne viruses by a highly efficient air sampler. ASM Biodefense Conference, Hyatt Regency Crystal City, Arlington, VA, USA, February 8 – 10, 2016.  Kenneth Rand, Maura Pieretti, Rodney Arcenas, Stacy Beal, Herbert Houck, Emma Boslet, **John Lednicky**. Semi-Quantitative Patient Data from a Multiplex Respiratory Viral Panel (RVP): Can We Learn Something from Population-Based Study? Annual University of Florida College of Medicine Celebration of Research, Monday, February 22, 2016, 5:30-8:30 pm Stephen C. O’Connell Center, UF.  Shannon Hentschel, Hao Chen, Julia Loeb, **John Lednicky**, and Tara Sabo-Attwood. Influenza infectivity modulation by carbon nanoparticles on sialic acid. EPI Research Day, Emerging Pathogens Institute, Gainesville, FL. 18 Feb. 2016.  Xiao Jiang, Christopher L. Cassano, **John Lednicky,** Chang-Yu Wu, and Z. Hugh Fan. Paper-based microfluidic devices for detecting RNA from flu virus. EPI Research Day, Emerging Pathogens Institute, Gainesville, FL. 18 Feb. 2016.  Sarah White, Wenjen Ma, Clinton McDaniel, Gregory Gray, and **John Lednicky.** Serologic evidence of exposure to influenza D virus among persons with occupational exposure to cattle. EPI Research Day, Emerging Pathogens Institute, Gainesville, FL. 18 Feb. 2016.  Hao Chen, Xiao Zheng, Justine Nicholas, Julia Loeb, Joseph H. Bisesi Jr., Sarah Robinson, **John Lednicky**, and Tara Sabo-Attwood. Single-walled carbon nanotubes suppress pulmonary immune response and increase infectivity on influenza virus exposed mice. EPI Research Day, Emerging Pathogens Institute, Gainesville, FL. 18 Feb. 2016.  Maha Elbadry, Valery Madsen Beau De Rochars, Massimilliano Tagliamonte, Mohammed Rashid, Jacques Boncy, Yves Jean Frantz Louis, J. Glenn Morris, Jr., **John Lednicky**, and Bernard Okech. Post-chikungunya fever epidemic cluster of dengue virus 1 infections among school children in Gressier Region, Ouest Department, Haiti. EPI Research Day, Emerging Pathogens Institute, Gainesville, FL. 18 Feb. 2016.  John Lednicky. Zika virus in Haiti in 2014: viral genomic and clinical data. Fourteenth Southeastern Regional Virology Conference (SERVC) 2016. Emory Conference Center, Atlanta, GA. 8 – 10 April, 2016.  Sarah White, Wenjen Ma, Clinton McDaniel, Gregory Gray, and **John Lednicky**. Serologic evidence of exposure to influenza D virus among persons with occupational exposure to cattle. PHHP Research Day, HPNP Reception Hall, Gainesville, FL. 13 Apr. 2016.  Maohua Pan, Arantzazu Eiguren-Fernandez, Nima Afshar-Mohajer, Susanne Hering, Chang-Yu Wu, **John Lednicky,** Hugh Fan, Hsin Hsieh.A novel sampler for virus aerosols through water-based condensation particle growth. 4th Workplace and Indoor Aerosols Conference, Barcelona, Spain. 20 – 22 April 2016.  **John Lednicky**, Maohua Pan, Julia Loeb, Hsin Hsieh, Arantzazu Eiguren-Fernandez, Susanne Hering, Z. Hugh Fan, Chang-Yu Wu. Highly Efficient Collection of Viable Influenza Virus A/Mexico/4108/2009 (pdmH1N1) Aerosols. 4th Workplace and Indoor Aerosols Conference, Barcelona, Spain. 20 – 22 April 2016.  Meeting abstract (presented in conference as a talk): Anderson BD, Ma M, Xia Y, Wang T, Shu B, **Lednicky JA,** Ma MJ, Lu J, Gray GC. Bioaerosol Sampling an Effective Approach to Studying Influenza A virus in Chinese Swine Farms. Options IX for the Control of Influenza, Chicago, IL. To be presented as an Oral Presentation at the International Society for Influenza and Other Respiratory Virus Diseases (ISIRV) 24 – 28 Aug. 2016.  Maohua Pan, Arantzazu Eiguren-Fernandez, Nima Afshar-Mohajer, Susanne Hering, Chang-Yu Wu, **John Lednicky,** Hugh Fan, Hsin Hsieh, and Patricia B. Keady. A Highly Efficient Sampler for Viable Virus Aerosols using Water-based Condensation Particle Growth. Biodefense World Summit, Baltimore, MD, June 27-30, 2016.  Meeting abstract (presented in conference as a talk): Haoran Yu, Chang-Yu Wu, Nima Afshar-Mohajer, **John A. Lednicky**, Z. Hugh Fan, Alexander Theodore and Liming Dong. Size Amplification and Preservation of the Viability of Aerosolized Virus during Collection by Batch Adiabatic-expansion for Size Intensification by Condensation (BASIC). AAAR 35th Annual Conference, 17 – 21 Oct. 2016.  Xiao Jiang, Maohua Pan, Susanne V. Hering, **John Lednicky**, Chang-Yu Wu, Z. Hugh Fan. Use of RNA Amplification and Electrophoresis for Studying Virus Aerosol Collection Efficiency and Their Comparison with Plaque Assays. AAAR 35th Annual Conference, 17 – 21 Oct. 2016.  Maohua Pan, Tania Bonny, Julia Loeb, Xiao Jiang, **John Lednicky,** Arantzazu Eiguren-Fernandez, Susanne Hering, Hugh Fan, Chang-Yu Wu. Collection of Viable Virus Aerosol in a Student Health Care Center through Water-Based Condensation Growth. AAAR 35th Annual Conference, 17 – 21 Oct. 2016.  Anderson BD. **Lednicky JA,** Gray GC. Bioaerosol Sampling In Swine Production Facilities: A Review Of The Literature. EcoHealth, 3 – 7 December 2016, Melbourne, Australia.  Maohua Pan, Tania Bonny, Julia Loeb, Xiao Jiang, John Lednicky, Arantzazu Eiguren-Fernandez, Susanne Hering, Hugh Fan, Chang-Yu Wu. Collection of airborne influenza virus in a student health care center through water-based condensation growth. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Tania S. Bonny, John P. Driver, Taylor Paisie, Marco Salemi, John Glenn Morris, Lisa A. Shender, Lisa Smith, Carolyn Enloe, Kevin Oxenrider, Jeffery A. Gore, Julia C. Loeb, Chang-Yu Wu, **John A. Lednicky**. Detection of alphacoronavirus vRNA in Brazilian free-tailed bats (*Tadarida brasiliensis*) from a colony in Florida, USA. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Sara Humes, Hao Chen, **John Lednicky**, Viet Dang, Nancy Denslow, and Tara Sabo-Attwood. Impacts of carbon nanotubes on lung cell lipidome and host immune responses following infection with pandemic influenza A virus. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Tania Bonny and **John Lednicky**. Isolation and identification of human coronavirus 229E (HCOV-229E) from frequently touched environmental surfaces in a classroom. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Xiao Jiang, Julia Loeb, **John Lednicky**, Chang-Yu Wu, and Hugh Fan. Paper-based microfluidic devices for detecting RNA from flu virus. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Hao Chen, Julia Loeb, Sara Humes, Sarah Robinson, **John Lednicky**, Tara Sabo-Attwood. Single-walled carbon nanotubes increase influenza A virus infectivity through oxidative stress in vitro. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Sarah White, **John Lednicky**, Valery Madsen Beau De Rochars, Maha Elbadry, Bernard Okech, and J. Glenn Morris. Detection of arbovirus co-infections of humans during a chikungunya virus outbreak, Haiti, 2014. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Sarah White, **John L**ednicky, James Dunford, and Bernard Okech. Detection of chikungunya-, dengue- and zika viruses in mosquitoes collected in Haiti, 2016. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Gabriela Blohm, **John Lednicky**, Alberto Paniz-Mondolfi, J. Glenn Morris, Jr., Marco Salemi, Julia Loeb, Sarah White, Taylor Paisie, and David Nolan. Evidence for transmission of zika virus from mother to baby by breast milk. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Gabriela Blohm**, John Lednicky**, Alberto Paniz-Mondolfi, Tania Bonny, Julia Loeb, Juliet Puliam, and J. Glenn Morris, Jr. Isolation of dengue 4 virus in clinical specimens from Venezuela during the outbreak of zika fever. Emerging Pathogens Institute Research Day, 23 Feb. 2017.  Mai-Juan Ma, MD, Guo-Lin Wang, PhD student, Benjamin Anderson, PhD, Zhen-Qiang Bi, MD, Bing Lu, MS, Xian-Jun Wang, MS, Chuang-Xin Wang, MS, Shan-Hui Chen, MS, Yan-Hua Qian, MPH, Shao-Xia Song, MS, Min Li, MS, John A. Lednicky, PhD, Teng Zhao, PhD student, Meng-Na Wu, MS student, Wu-Chun Cao, PhD and Gregory Gray, MD, MPH. Evidence for Cross-species Influenza A Virus Transmission within Swine Farms, China. Abstract # 64165, Submitted to IDWeek 2017. 12 May 2017.  Sarah White, John Lednicky, James Dunford, Bernard Okech and J. Glenn Morris Jr., University of Florida, Gainesville, FL, Navy and Marine Corps Public Health Center, Portsmouth, VA ESA Section: Medical, Urban, and Veterinary Entomology (MUVE) - Regular, Non-Competition Poster: Detection of Chikungunya-, Dengue- and Zika viruses in mosquitoes collected in Haiti, 2016. Submission # 123758. 13 May 2017. |

**Recent talks**

A nose-only inhalation exposure system for studies of aerosolized pathogens in small animals. College of Veterinary Medicine, University of Florida, Gainesville, Florida, 27 Sept. 2011.

Refinement of Virus Detection and Isolation Methodologies for Aerobiology Studies. Viral Surrogate Pathogen Workshop, 16 May 2012, Naval Research Laboratory, Washington, DC.

Recent studies regarding the aerobiology of influenza and other respiratory viruses. Public Health Seminar Series, College of Public Health and Health Professions. University of Florida, 18 Feb. 2013.

Collection of viable airborne viruses using air samplers. Biomedical Education Seminar Series. Burnett School of Biomedical Sciences, University of Central Florida, 7 February 2014.

**Keynote Address:** One Health – A multidisciplinaryapproach to monitor and control public health threats. BioKansas; Fifth Annual One Health Summit. March 6, 2014, Sporting Park, Kansas City, Kansas.

Experiments on the collection of airborne viruses in public airspaces. Emerging Pathogens Institute, University of Florida, Gainesville, Florida, 14 March 2014.

The Ebola Outbreak in West Africa: Understanding the Virus and the Public Health Challenges It Poses. John Lednicky (Part 1: Virology and Clinical Aspects) and Paul Psychas (Part 2: Epidemiology and Public Health). Emerging Pathogens Institute, University of Florida, Gainesville, Florida, 27 August 2014.

Therapeutic Potential of a-Neurotoxoids. Three-part presentation by Jay Yourist (Part 1), Patrick Corsino (Part 2), and **John Lednicky** (Part 3). Office of Biodefense, Research Resources and Translational Research, Division of Microbiology and Infectious Diseases National Institute of Allergy and Infectious Diseases, National Institute of Health, Rockville, Maryland, 11 Sept. 2014.

Ebola: The Disease, the Virus, the Current Outbreak, and Infection Control Practices. Veterans Health Administration's National Center for Occupational Health and Infection Control, Commerce Building, Gainesville, Florida, 8 Oct. 2014.

Ebola 101. Presented at the UF Mini Medical School Symposium. Cancer Genetics Research Complex, Auditorium (Room 101), UF, Gainesville, Florida, 21 Nov. 2014.

Improvements in air sampling methodologies for studies of airborne respiratory pathogens. John Lednicky. 3rd Global Centre for Mass Gathering Medicine Scientific Advisory Board Meeting. Radisson Blu Hotel, Riyadh, Saudi Arabia. 29 March, 2015.

Some recent surprises regarding tick-borne viruses. John Lednicky. 3rd Global Centre for Mass Gathering Medicine Scientific Advisory Board Meeting. Radisson Blu Hotel, Riyadh, Saudi Arabia. 30 March, 2015.

Zika virus in Haiti in 2014: viral genomic and clinical data. Fourteenth Southeastern Regional Virology Conference (SERVC) 2016. Emory Conference Center, Atlanta, GA. 8 – 10 April, 2016.

A new approach to sampling infectious bioaerosols. Invited (closed door) presentation. Talk given in 3 parts: by Pat Keady, AEROSOL DEVICES INC., Fort Collins, CO, and C.Y. WU and **John Lednicky**, Univ. FL, 22 Sept. 2016, Dept. Homeland Security Headquarters, Washington, DC.

*Zika virus* in the Americas: why were we caught off-guard? Gainesville Rotary Club, held at Paramount Grill, Gainesville, Florida, 8 Dec. 2016.

*Zika virus*: Recent findings. Seminar series, Dept. of Environmental and Global Health, Gainesville, FL, 31 Jan. 2017.

Major Surprises Arising from Improved Virus Detection and Isolation Methods. 5 May 2017. Emerging Pathogens Institute, University of Florida.

**Recent short oral presentations**

Detection, Collection, and Modeling of Viable Airborne Viruses: Presentation for: James F Cummings, COL USARMY MEDCOM AFHSC (US). Presentation at University of Florida Emerging Pathogens Institute, May 9, 2013.

Aerobiology, Respiratory Pathogens, and Virus Discovery: Presentation for Department of Defense (Capt. Lax and other representatives) at Nuovo Biologicals, Davie, Florida. 22 Aug. 2013.

Collection of viable airborne viruses using air samplers. Presentation for Project HOPE representatives. EPI, 4th floor conference room, 7 Feb, 2014.

Aerobiology, respiratory pathogens, and virus discovery. 2014 Spring HHMI Science for Life Seminar Class, University of Florida –Gainesville, 13 February 2014.

MPH Common Reader Day (Group Discussion of Book: Spillover by David Quammen). PHHP MPH students, 16 Sept. 2015, Univ. Florida, Gainesville, FL.

Isolation of *Zika virus* from three different plasma specimens collected in December 2014 from Haitian children with febrile illness and complete genomic sequence analysis of one of the virus isolates. 2 Feb. 2016, Emerging Pathogens Institute, Univ. Florida, Gainesville, FL.

*Zika virus* explained. Presentation by Drs. Amy Vittor (Where did Zika virus come from?), Michael Weiss (What is microcephaly?), John Lednicky (*Zika virus*: some basic information), Jorge Rey (Mosquito vectors in Florida), Danielle Stanek (Current status), and Glenn Morris (Summary). Presented to Florida Legislature (teleconference/slide presentation). Emerging Pathogens Institute, Univ. Florida, Gainesville, FL. 19 Feb. 2016.

*Zika virus* in Haiti in 2014: virology findings. Private presentation (evening, 9 April 2016) to virology group attending Fourteenth Southeastern Regional Virology Conference (SERVC) 2016. Emory Conference Center, Atlanta, GA.

*Zika virus*; Recent findings. Private presentation (evening, 11 April 2016) to arbovirologists. Gainesville, FL.

*Zika virus* in Haiti in 2014: Summary of findings. Presented to Dr. James Dunford, entomologist, US Navy Environmental Preventive Medicine Unit 2 (NEPMU-2). Lednicky office, Dept. Conference Room, and Lednicky laboratory. 11 Apr. 2016.

Zika. Talk given at the Department of Health, Alachua County, 224 SE 24th Street

Gainesville, FL 32641. 11 May 2016.

Virus discovery and characterization. Presentation to Emerging Pathogens Institute External and Internal Advisory Board, UF Cypress Lodge, Lake Wauberg, Micanopy, FL. 24 Feb. 2017.

*Zika virus* work at the Emerging Pathogens Institute. Discussion with State of Florida Education Appropriations Committee, Florida State Capitol Building, Tallahassee, Florida, 20 March 2017.

*Zika virus* – Lednicky Laboratory. Presentation to Florida Lieutenant Governor Carlos Lopez-Cantera. Emerging Pathogens Institute, UF, 23 March 2017.

Capacity Building: Establishing the CHeRI Cervid Virus Research Laboratory. 1st Annual Cervidae Health Research Initiative. Harn Museum of Art Auditorium, UF Campus, Gainesville, Florida 29 March 2017.

**Recent Grant Proposal Review Panel Participation**

Defense Threat Reduction Agency's Chemical Biological Technologies (DTRA) Directorate FY12/13 Service Call for proposal number CBCALL12-DIAGB1-1-0105. 1/7/2011 – 2/11/2011

CDC Special Emphasis Panel – FOA CK11-006; Combining subjective and objective methods for quantifying contact rates and mixing patters in school-aged children. 4/13/2011 – 5/3/2011.

NIH NIAID Special Emphasis Panel for Phased Innovation Awards on Partnerships for Interventions to Treat Chronic, Persistent and Latent Infections. 10/17/2012 – 12/12/2012. ZAI1 JKB-M J4; Jane Battles, SRO R1/R33 RFA-A1-12-020

CDC Special Emphasis Panel for HJFMRI Public Health Research in Kenya, ZGHGH16-006 KENYA, Panel A. 17 May 2016.

CDC Special Emphasis Panel for Household Transmission of Influenza Viruses in the Community, RFA-IP-17-001, May 5, 2017.

**Primary Isolation and Characterization of SV40 Viruses**

SV40-strain CPC/MEN. Described in Lednicky et al., Virol. 212, 710-717 (1995) and in Stewart et al., J. NeuroVirol. 4, 182-193 (1998). Plasmid clone deposited in ATCC (ATCC # VRMC-4).

SV40-strain K661. Described in: Lednicky et al., J. Virol. 72, 3980-3990 (1998). Plasmid clone deposited in ATCC (ATCC # 87722).

SV40-strain T302. Described in: Lednicky et al., J. Virol. 72, 3980-3990 (1998) (ATCC # VRMC-14).

SV40-6593-2. Described in: Lednicky et al., J. Virol. 72, 3980-3990 (1998).

SV40-Baylor-1 (SV40-B1). Described in: Lednicky, J. A. and J. S. Butel, J. Gen. Virol. 78, 1697-1705 (1997). Plasmid clone deposited in ATCC (ATCC # VRMC-2).

SV40-Baylor-3 (SV40-B3). (Unpublished)

SV40-CAL-1. Described in: Virus Genes 29, 183-190 (2004).

Numerous additional plasmid clones (containing full-length virus genomes) have been deposited at ATCC.

**Primary Isolation and Characterization of Miscellaneous Viruses, many of which are Difficult to Isolate *In-Vitro* (partial list)**

Canine coronavirus

Canine distemper American type 2 (numerous strains from dogs and wildlife)

Canine influenza virus

*Human coronavirus NL63*

*Human parainfluenza virus 4A*

*Human parainfluenza virus 4B*

JCV-JAL-1

*Kilham rat virus*-JAL-1

Sea lion adenovirus

Cervid poxvirus

Epizootic hemorrhagic disease virus 1, 2, and 6

Zika, Dengue 1-4, Chikungunya viruses

**Molecular Detection of Non-Culturable Viruses**

*Human polyomavirus 9* (new variant), tentatively designated type C (partial clone available)

*Human rhinovirus 51-C* strain JAL (unique and optimal reverse genetics system clone has been

prepared for this virus)

**GenBank Submissions**

Dr. Lednicky served as primary author on >200 entries for a wide variety of viruses, from canine distemper to human parainfluenza. He is a co-author of many other entries, including the full genomic sequences of *West Nile Virus*, *Eastern equine encephalitis virus*, *Japanese encephalitis virus* and more recently, *Human parainfluenza 4B virus, Human coronavirus NL63, Human rhinovirus 51-C.*